# Environmental enrichment increases brain volume in snakes

Gokulan Nagabaskaran, Vijay Moonilal, Morgan Skinner, Noam Miller

Department of Psychology, Wilfrid Laurier University

This is a preprint of an article to appear in The Journal of Comparative Neurology. A link to the final version will be added here when available.

## Abstract

The effects of environmental enrichment have been well documented in mammals and birds, but less work has focused on reptiles. Because snakes are common in captivity, both as pets and in research/commercial facilities, it is critical to explore how they react to standard captive housing. Here, we examined the effects of environmental enrichment on brain development in a popular pet snake species, the western hognose snake (*Heterodon nasicus*). Hognose snakes (n=15) were individually housed for one year in either enriched or standard environments, before their brains were harvested and imaged using MRI. We found that enriched snakes had significantly larger brain volumes compared to standard snakes, most prominently in posterior brain regions. Additionally, we observed sex-specific brain investments: as snakes grew larger, males displayed relatively larger cerebral hemispheres and females displayed larger posterior brain regions. These results suggest that environmental enrichment is critical to encouraging healthy brain development in snakes, and that snake brain plasticity is very similar to that observed in mammals and birds.

Keywords: snake, reptile, environmental enrichment, brain, MRI, reptile welfare, 3D Slicer, RStudio

## Introduction

Many animals in captivity are housed under less than ideal conditions. Captive environments may lack critical aspects of a species' natural habitat and may not allow animals to engage in natural behaviors important to their health and wellbeing (Young et al., 2020). Deficiencies in captive environments can be identified by improvements in cognition and health – as well as behavioral changes – that emerge when environments are improved or enriched (Olsson and Dahlborn, 2002; Tahamtani et al., 2018; Toli et al., 2016). Environmental enrichment (EE) can be defined as changing a captive animal's environment in a beneficial manner that encourages natural behaviours (Widenmayer, 1996; Young et al., 2020) which may reduce stress and improve health (Leal-Galicia et al., 2008; Coulton et al., 1997). EE can take many forms and should be tailored to the ecology of the animal (Young et al., 2020). One common method of EE is to increase the complexity of the captive environment using physical structures (Depasquale et al., 2016) that allow animals to engage in natural behaviours such as burrowing, nesting, or climbing (Van de Weerd et al., 1996; Hoehfurtner et al., 2021).

EE can lead to cognitive benefits, such as improved spatial learning, increased exploration, increased memory retention, and decreased anxiety (Jones and Waddington, 1992; Leal-Galicia et al., 2008; Depasquale et al., 2016). Such cognitive changes are inevitably linked to changes in the brain, with studies observing increases in brain size (Cummins et al., 1973; Scotto Lomassese et al., 2000), increased neurogenesis (Depasquale et al., 2016; Leal-Galicia et al., 2008; Segovia et al., 2006), and effects on disease progression (Wolf et al., 2006). Increases in brain size as a result of enrichment have been observed in various animal models and are sometimes age-dependent (Cummins et al., 1973; Fong et al., 2019; Naslund et al., 2012).

In fish, enrichment primarily affects growth in the hippocampal formation (Depasquale et al., 2016), but inconsistent results have been found in other taxa. When stones were added as enrichment to juvenile steelhead salmon tanks (*Oncorhynchus mykiss*), their cerebellums grew larger compared to fish in barren

tanks (Kihslinger and Nevitt, 2006). However, coho salmon (*Oncorhynchus kisutch*) reared in simplistic hatchery environments grew larger brains overall than fish raised in a complex stream-like environment (Kotrschal et al., 2012). Other studies have also produced mixed, little, or no effects as a result of enrichment across various species (Toli et al., 2017; Depasquale et al., 2016; Kihslinger and Nevitt, 2006).

Little work has been done on EE in reptiles, compared to mammals and birds, even though reptiles engage in complex behaviours that include social communication, social learning, parental care, and play (Font et al., 2023). The few reptilian studies conducted have been inconclusive, with some showing a positive effect of enrichment and others not, depending on factors such as ontogeny and species-specific needs (Burghardt and Layne, 1995; Nagabaskaran, 2020). Rat snakes (Elaphe obsoleta) exposed to EE displayed quicker habituation rates and performed better in goal oriented tasks compared to unenriched snakes (Almli and Burghardt, 2006). Enriched corn snakes (Pantherophis guttatus) displayed discrimination of a familiar handler based on odor cues and familiar objects that lacked odor cues, while snakes housed without enrichment did not (Nagabaskaran et al., 2021; Nagabaskaran, 2020). Ball pythons (Python regius) and Madagascar giant hognose snakes (Leioheterodon madagascariensis) display significantly fewer stress-induced abnormal behaviours in enriched environments (Hollandt et al., 2021; Spain et al., 2020), and leopard geckos (Eublepharis macularius) display high levels of interest and motivation for enrichment after being exposed to barren environments (Zielikski, 2023). Moreover, both western hognose snakes (*Heterodon nasicus*) and corn snakes show a clear preference for EE when given a choice between enriched and unenriched environments (Nagabaskaran et al., 2022; Hoehfurtner et al., 2021). These studies used naturalistic EE that allowed for climbing, burrowing and access to multiple shelters that varied in elevation and humidity. Such components of enrichment, that reflect the species' natural environment, should be used widely with all captive reptiles to improve reptilian health and welfare (Warwick & Steedman, 2023), as a survey of 675 snakes worldwide indicated that snakes with larger enclosures and enrichment stimuli displayed fewer abnormal clinical signs (Cargill et al., 2022).

In contrast, a few studies have found no significant effects of enrichment on reptiles. For example, arboreal fence lizards (*Sceloporus undulatus*) showed no change in either natural behaviours or stress hormone levels when provided with a fourteen centimeter high climbing enrichment (Rosier and Langkilde, 2011), which may have simply been inadequate given the much larger heights these lizards naturally climb to (Kennedy, 1958). Similarly, green anoles (*Anolis carolinensis*) and corn snakes displayed no significant changes in preference, behaviour, or physiological markers when exposed to environmental enrichment (Borgmans et al., 2018; Evans, 2011). Additionally, box turtles (*Terrapene carolina*) displayed some significant changes when exposed to enrichment but these effects were transient across multiple timepoints (Tetzlaff et al., 2019). Given this inconsistency in the data on enrichment in reptiles, further investigation is clearly required.

Even less work has been done on the effects of enrichment on the brains of reptiles. A territorial morph of side-blotched lizards (*Uta stansburiana*) displayed neurogenesis when given EE in the form of a larger enclosure, though non-territorial morphs did not (LaDage et al., 2013), and there is a mention of enriched monitor lizards displaying a trend for larger forebrains in an unpublished note (in Burghardt, 2013). These results hint that reptilian brains are plastic, like those of other taxa, and that they may be affected by environmental factors.

To better understand the effects of enrichment on reptile brains, we assessed whether and how providing EE affected brain volume in juvenile western hognose snakes. This species is semi-fossorial and is known for its unique shovel-like nose that it uses to dig underground shelters. They are opportunistic feeders that prey on both aquatic and terrestrial organisms (Averill-Murray, 2006). We exposed half our snakes to species-specific enrichment and housed the remaining subjects in a relatively barren rack system, similar to those commonly used by breeders and pet owners (Figure 1B). After living in their respective treatments for approximately one year, we obtained MRI scans of all the snakes' brains. Paralleling findings in other taxa, we hypothesized that exposure to EE would increase brain volumes in developing snakes.

## Methods

#### Animals & Husbandry

15 captive-bred western hognose snakes (five males) were acquired from a local breeder. The snakes weighed an average of  $4.3 \pm 3.2$  g at the start of the study and were 2-3 months old. They were housed individually in either enriched or standard enclosures (see below) for the duration of the study (~ 1 year). The snakes were fed two pieces of human-grade defrosted salmon (Great Value) dusted with reptile calcium supplement (Zilla) weekly on the same day, from within individual feeding boxes (11 x 15 x 4 cm). Each piece of salmon was slightly larger than the snake's head. In both conditions, snakes were given fresh water daily. Any feces or sheds found were removed during daily checks, and when snakes in the standard housing condition defecated, the entire paper-towel flooring was replaced.

## Housing Conditions

Three males and five females were housed in the enriched condition, and the remaining two males and five females were housed in the standard condition. The enriched enclosures (46 x 56 x 30 cm) were constructed of PVC with a sliding glass door (43 x 12 cm) in the front. Enrichment was chosen based on preliminary preference testing (Nagabaskaran et al., 2022) and consisted of approximately 5 cm deep loose coconut husk substrate (Zoo Med Eco), a single black plastic shelter (Cornel's World; 14 x 10 x 5 cm), a large water dish (27 x 15 x 6 cm), a sand dish for burrowing, plastic vine (Cornel's World), a damp shelter (10 x 7 cm), and straw balls (5 cm diameter) for rooting behaviour (see Figure 1B). Enclosures included LED lighting (at 2700 Kelvin) on the ceiling with a 12:12 hr cycle (lights on at 8:30 am), and heat was provided via thermostat-controlled heat-tape (THGHeat; Spyder Electronics HerpStat) under one corner of the enclosure. This created a heat gradient within the enclosure ranging from 23 to 32 °C. Heat tape was used as it maintains constant temperatures better than heat lamps and mimics commonly used heating setups by pet owners.

Snakes in the standard condition were housed in drawer-sized boxes ( $24 \times 38 \times 8 \text{ cm}$ ) placed within bare enclosures similar in size to the enriched enclosure. The same light cycle and heating were provided. The enclosures had paper towel sheets as bedding, a single black shelter (Cornel's World;  $14 \times 10 \times 5 \text{ cm}$ ) and a small water dish in the form of a plastic sauce cup ( $6 \times 3 \text{ cm}$ ).

## Perfusion/MRI Methodology

Snake brains were prepared and scanned using well established procedures for imaging small animal brains, for which euthanasia is required (Spring et al., 2007; Cahill et al., 2012; Ellegood et al., 2014). Snakes were anesthetized with an overdose of sodium pentobarbital (100 mg/kg body weight) injected intracardially, based on weights taken immediately prior to sedation. Snakes were checked carefully for reactivity after dosing to ensure proper sedation via strong tail pinches and assessment of muscle tone, and only progressed to perfusion after body muscle contractions and tongue movements in reaction to pinching had ceased. They were perfused transcardially with 100 mL of 0.1 M Phosphate Buffered Solution (PBS; pH 7.3) containing 2 mM ProHance (a contrast agent; gadoteridol, Bracco Diagnostics Inc., Princeton, NJ) followed by 100 mL of 4% paraformaldehyde with 2 mM ProHance. This was necessary to allow the contrast agent to perfuse the brain. After perfusion, the snakes were immediately decapitated, and the tissue surrounding the skull was removed. The skulls containing the brains were placed in 4% paraformaldehyde with 2 mM ProHance at 4 °C for approximately 12 hours immediately after surrounding tissue was removed. The skulls were then transferred to a 0.1 M PBS solution containing 2 mM ProHance and 0.02% sodium azide until they were scanned.

Brain images within the skulls were acquired using a 3 Tesla MRI scanner (Siemens PrismaFit) at The Center for Phenogenomics (TCP) in Toronto, Canada. Images were T2 weighted. All images were processed using 3D Slicer 5.2.2 (Federov et al., 2012; <u>https://www.slicer.org</u>) by a researcher blind to the identities of the snakes and their housing condition (VM). Segmentation was performed using 3D Slicer's automated *Grow from seeds* tool, combined with manual adjustment when necessary. Complete brains

were further segmented into three subsections, based on easily identifiable morphological features of the snake brain. This involved segmentation at the junction of the olfactory bulbs and the cerebral hemispheres, as well as at the sulcus where the forebrain and midbrain meet (Figure 3A). These three subsections are hereafter referred to as the olfactory bulbs (OB; including the main olfactory bulb, accessory olfactory bulb, and the olfactory tract), the cerebral hemispheres (CH, approximately overlapping with the pallium), and the posterior brain (PB; the midbrain and hindbrain, including the optic tectum, cerebellum and brainstem). Overall brain volumes (in mm<sup>3</sup>), as well as the volumes of each subsection, were calculated using 3D Slicer's *segment statistics* module. Volumes for each subsection were converted to proportions of total brain volume.

#### Statistical analysis

Analyses were performed in R (version 4.2.1, R Core Team, 2022). We modeled the factors important for predicting brain volume by progressively adding weight, sex, a weight by sex interaction, and housing condition to an analysis of variance. We compared the resulting models by their AIC values and dropped factors that did not improve the model fit. A model was considered to be a significantly better fit if it improved the AIC by a value of 2 or more (Cavanaugh and Neath, 2019). As brain volumes were strongly correlated with snakes' overall weight (see Results), we included weight as a covariate in all models.

#### Results

Larger snakes had larger brain volumes (Figure 1A; F(1,12) = 22.54, p < 0.001), and snakes in the enriched housing condition had larger brains than controls (Figure 1B; F(1,12) = 5.34, p = 0.039). Further analysis found that most of the increase in volume for snakes in the enriched condition occurred in the posterior brain (Figure 2E, H; F(1,12) = 6.73, p = 0.025; Enriched mean =  $31.81 \pm 2.91$ , Standard mean =  $30.82 \pm 3.31$ ), with no significant effect found in the cerebral hemispheres (Figure 2D, G; F(1,12) = 1.76, p = 0.209; Enriched mean =  $23.86 \pm 2.12$ , Standard mean =  $23.46 \pm 1.60$ ) or the olfactory bulbs (Figure

2F, I; F(1,12) = 1.43, p = 0.256; Enriched mean = 5.66 ± 0.71, Standard mean = 5.66 ± 1.14). Larger snakes had larger brains in all three subregions (all F's > 7.00, all p's < 0.05).



**Figure 1**: Brain volume as a function of weight, sex, and enrichment. **A**: brain volume in mm<sup>3</sup> as a function of snake weight in g for male (M, square) and female (F, circle) snakes in the enriched (green) and standard (orange) conditions. The black diagonal line shows a linear regression on the data. **B**: brain volumes for the enriched (green) and standard (orange) snakes, controlled for snake weight; error bars show  $\pm$  SEM. The insets show photographs of the enriched and standard environments.

We also examined the volumes of the subregions as a proportion of overall brain size. We found an interaction between weight and sex in both the cerebral hemispheres and the posterior brain (CH: F(1,11) = 7.08, p = 0.022; PB: F(1,11) = 7.99, p = 0.017), but these interactions were in opposite directions: as a proportion of total brain volume, larger male snakes tended to have larger cerebral hemispheres, while the opposite pattern occurred in females (Figure 2A). In the posterior brain, larger females tended to have larger volumes, whereas larger males tended to have less volume (Figure 2B). For both these brain regions, there were no main effects of weight (CH: F(1,11) = 3.49, p = 0.089; PB: F(1,11) = 0.27, p = 0.614), or sex (CH: F(1,11) = 0.06, p = 0.806; PB: F(1,11) = 0.06, p = 0.819). For the olfactory bulbs, we did not find an interaction, but larger snakes had disproportionately larger olfactory bulbs (Figure 2C; F(1,13) = 6.2, p = 0.027). In summary, enrichment tended to increase the brain volume of the snakes, especially in posterior regions of the brain. In addition, as female snakes grew, they tended to invest more volume into posterior brain areas and their olfactory bulbs. In contrast, as males grew, they invested more

volume in the cerebral hemispheres and olfactory bulbs. There was no effect of housing condition on the relative volume of any brain region.



**Figure 2**: Relative (A-C) and absolute (D-I) brain area volumes as a function of weight, sex, and enrichment. A,D,G: forebrain; B,E,H: mid/hindbrain; C,F,I: olfactory bulb. Panels A-C show the proportion of total brain volume consisting of that brain area as a function of snake weight in g for male (blue) and female (red) snakes in the enriched (circle) and standard (square) conditions. The blue and red lines in each panel show linear regressions. Panels D-F show absolute brain volumes (in mm<sup>3</sup>) of male snakes in the enriched (circle) and standard (square) conditions, and panels G-I show the same data for female snakes.



**Figure 3**: brain segmentation. **A**: 3D view of a sample segmented brain; **B**: sample coronal slice; **C**: sample sagittal slice; **D**: sample horizontal slice. In all panels, olfactory bulbs are shaded brown, cerebral hemispheres in purple, and posterior brain in green. The gray areas in panels B-D are non-brain tissues. All images generated by *3D Slicer* (v. 5.2.2).

## Discussion

We investigated the effects of naturalistic environmental enrichment (EE) on brain volume in juvenile western hognose snakes. We found that exposure to physical EE during the first year of life significantly increased total brain volume, particularly in posterior brain regions (midbrain and hindbrain). Our findings add to growing evidence that reptiles respond to EE in a similar manner to other well studied animal models (Cummins et al., 1973; Scotto Lomassese et al., 2000; Naslund et al., 2012), in contrast to the common misconception that reptiles have a limited behavioral repertoire and are tolerant of minimalistic housing conditions (Warwick, 1990; Case et al., 2005). As undisturbed natural environments are likely to be more complex and challenging than captive conditions, it may be more accurate to state that standard reptile housing conditions stunt brain growth and have negative effects on cognition and welfare. This is because captive environments are generally representative of what humans expect animals need, often leading to "controlled deprivation" (Burghardt, 1996), in which captive environments lack many components necessary for healthy growth (Mendyk & Augustine, 2023; Mendyk, 2018).

The EE that we implemented took the form of naturalistic stimuli (substrate, climbing vines, large water dishes) that allowed snakes to engage in behaviours like climbing, burrowing, and swimming, thus encouraging various forms of locomotion. The enriched enclosures were also larger, providing more space for locomotion (Figure 1). Several studies observing the effects of enclosure size on reptiles have found that more space is generally positive. Corn snakes that were kept in enclosures large enough to completely stretch out were found to be more active and showed a significant preference for larger enclosures when allowed to actively choose (Hoehfurtner et al., 2021). Eastern blue-tongued lizards (*Tiliqua scinoides*) utilized extra enclosure space when provided by engaging in more activity throughout the enrichment duration, encouraging healthy weight management (Philips et al., 2023). Various species of turtles also display an increase in the diversity of behaviours when exposed to EE in the form of larger space and more stimuli (Turner et al., 2022). The increased level of exercise available to snakes in the enriched condition may have been partially responsible for their larger brain volume, consistent with

findings in other taxa (Cummins et al., 1973; Scotto Lomassese et al., 2000; Fong et al., 2019; Naslund et al., 2012).

There is very little literature on anatomical effects of EE in reptiles. Juvenile male side-blotched lizards displayed increased neurogenesis in the medial cortex when exposed to larger enclosures for 5 months, but the effect was confined to territorial males; no overall differences in brain volume were found (LaDage et al., 2013). This contrasts with research demonstrating larger overall brain volumes in wild territorial male side-blotched lizards compared to non-territorial males (LaDage et al., 2009). LaDage and colleagues (2013) explained these differences by suggesting that their EE lacked the complexity of the lizards' natural environment (LaDage et al., 2013). In line with this suggestion, our results indicate that the size and complexity of the environment is important for effects on brain volume in reptiles. Based on our findings it is hard to discern whether this difference in brain size is a result of increased space or increased diversity of enrichment stimuli. Future research should determine the exact combination of enriching factors required for brain volume increases as well as how long animals need to spend under these conditions and whether there are critical periods for the effects.

The overall increase in brain volume we observed in enriched hognose snakes occurred primarily in the posterior brain, made up of both the midbrain and the hindbrain and containing the optic tectum (midbrain), medulla oblongata and cerebellum (hindbrain). Similar results have been reported in Atlantic salmon (Naslund et al., 2012). The snake cerebellum has been suggested to be extremely sensitive to locomotory behaviours, and its shape may depend on the types of locomotion snakes engage in (Macri et al., 2023). The significant increase in the size of the posterior brain in our enriched snakes may be partly explained by increased demand on their cerebellums. In reptiles, the optic tectum is involved in visual and somatosensory processing (Catania, et al., 2010), and the medulla oblongata in sensory and motor processing (Senn et al., 1970). As the enrichment we provided was both visually and tactilely more complex than the unenriched cages, it is possible that increased sensory complexity drove some of the increases in the size of these sensory-motor regions.

In addition to EE-driven differences in brain size, we also observed differences between sexes. As they grew larger, female juvenile snakes displayed a larger posterior brain region while males displayed larger cerebral hemispheres, regardless of housing treatment. We note that, though our snakes were all approximately the same age, weight is a more important factor in determining maturity in snakes (Shine & Charnov, 1992; Feldman & Meiri, 2012). Sexual dimorphism in brain sizes is not uncommon and is evident in mammals, birds and fish (Toli et al., 2016; Gittleman, 1994; Garamszegi et al., 2005). In general, male and female snakes face similar environmental challenges, but they differ in breeding strategies and requirements which may explain the differences we observed (Garamszegi et al., 2005).

In conclusion, we found that physical enrichment increased overall brain size in hognose snakes – as it does in mammals, birds, and fish – mostly in posterior brain regions. We also observed sex-specific investments in different subregions of the brain. Our results, combined with earlier work on preference for enrichment (Nagabaskaran et al., 2022; Hoehfurtner et al., 2021) and its cognitive benefits (Nagabaskaran et al., 2021; Nagabaskaran, 2020) in snakes, strongly suggest that snakes (and probably other reptiles) react to the complexity and diversity of behavioral opportunities in their environments in much the same way that mammals and birds do. We should therefore consider their needs in captivity in the same way as we do members of those other taxa.

## References

- Almli LM, Burghardt GM (2006). Environmental enrichment alters the behavioural profile of ratsnakes (*Elaphe*). *Journal of Applied Animal Welfare Science*, *9*:85–109.
- Averill-Murray RC (2006). Natural history of the western hog-nosed snake (Heterodon nasicus) with notes on envenomation. *Sonoran Herpetologist, 19*:98-101.
- Borgmans G, Palme R, Sannen A, Vervaecke H, Van Damme R (2018). The effect of environmental provisioning on stress levels in captive green anole (*Anolis carolinensis*). *Animal Welfare*, 27:35-46.
- Burghardt GM, Layne DG (1995). Effects of ontogenetic processes and rearing conditions. In *Health and welfare of captive reptiles* (pp. 165-185). Dordrecht: Springer Netherlands.
- Burghardt GM (1996). Environmental enrichment or controlled deprivation? In GM Burghardt, GM
   Bielitski, JT Boyce, JRDO Schaefer (Eds.), *The Well-Being of Animals in Zoo and Aquarium Sponsored Research* (pp. 91-101), Scientists Center for Animal Welfare.
- Burghardt GM (2013). Environmental enrichment and cognitive complexity in reptiles and amphibians: Concepts, review, and implications for captive populations. *Applied Animal Behaviour Science*, 147:286-298.
- Cahill LS, Laliberté C, Ellegood J, Spring S, Gleave JA, Van Eede MC, Lerch JP, Henkelman RM (2012). Preparation of fixed mouse brains for MRI. *NeuroImage*, 60:933–939.
- Cargill BM, Benato L, Rooney NJ (2022). A survey exploring the impact of housing and husbandry on pet snake welfare. *Animal Welfare*, *31*:193-208.
- Case BC, Lewbart GA, Doerr PD (2005). The physiological and behavioural impacts of the preference for an enriched environment in the eastern box turtle (*Terrapene carolina carolina*). *Applied Animal Behaviour Science*, 92:353–365.
- Catania KC, Leitch DB, Gauthier D (2010). Function of the appendages in tentacled snakes (*Erpeton tentaculatus*). Journal of Experimental Biology, 213: 359-367.

- Cavanaugh JE, Neath AA (2019). The Akaike information criterion: Background, derivation, properties, application, interpretation, and refinements. *WIREs Computational Statistics*, 11:e1460.
- Cummins RA, Walsh RN, Budtz-Olsen OE, Konstantinos T, Horsfall CR (1973). Environmentallyinduced changes in the brains of elderly rats. *Nature*, 243:516–518.
- DePasquale C, Neuberger T, Hirrlinger AM, Braithwaite VA (2016). The influence of complex and threatening environments in early life on brain size and behaviour. *Proceedings of the Royal Society B*, 283:20152564.
- Ellegood J, Anagnostou E, Babineau BA, Crawley JN, Lin L, Genestine M, et al. (2014). Clustering autism: using neuroanatomical differences in 26 mouse models to gain insight into the heterogeneity. *Molecular Psychiatry*, 20:118–125.
- Evans SL (2011). Effect of environmental enrichment on leukocytes and housing preference in captive red cornsnakes, *Pantherophis guttatus* (PhD Dissertation). Emporia State University.
- Feldman A, Meiri S. (2012). Length-mass allometry in snakes. *Biological Journal of the Linnean Society/Biological Journal of the Linnean Society*, *108*(1): 161–172.
- Fong S, Beuchel SD, Boussard A, Kotrschal A, Kolm N (2019). Plastic changes in brain morphology in relation to learning and environmental enrichment in the guppy (*Poecilia reticulata*). Journal of Experimental Biology, 222:jeb200402.
- Font E, Burghardt GM, Leal M (2023). Brains, behaviour, and cognition: Multiple misconceptions. In *Health and welfare of captive reptiles* (pp. 211-238). Cham: Springer International Publishing.
- Garamszegi LZ, Eens M, Erritzøe J, Møller AP (2005). Sperm competition and sexually size dimorphic brains in birds. *Proceedings of the Royal Society B*, 272:159-166.
- Gittleman JL (1994). Female brain size and parental care in carnivores. *Proceedings of the National Academy of Sciences*, 91:5495-5497.
- Hoehfurtner T, Wilkinson A, Nagabaskaran G, Burman OHP (2021). Does the provision of environmental enrichment affect the behaviour and welfare of captive snakes? *Applied Animal Behavior Science*, 239:105234.

- Hoehfurtner T, Wilkinson A, Walker M, Burman OH (2021). Does enclosure size influence the behaviour
  & welfare of captive snakes (*Pantherophis guttatus*)? *Applied Animal Behaviour Science*, 243:105435.
- Hollandt T, Baur M, Wöhr AC (2021). Animal-appropriate housing of ball pythons (*Python regius*) Behavior-based evaluation of two types of housing systems. *PLoS One*, *16*:e0247082.
- Jones RB, Waddington D (1992). Modification of fear in domestic chicks, *Gallus gallus domesticus*, via regular handling and early environmental enrichment. *Animal Behaviour*, 43:1021-1033.
- Kennedy JP (1958). Sleeping habits of the eastern fence lizard, *Sceloporus undulatus hyacinthinus* (Sauria, Iguanidae). *The Southwestern Naturalist*, 1:90–93.
- Kihslinger R, Nevitt GA (2006). Early rearing environment impacts cerebellar growth in juvenile salmon. Journal of Experimental Biology, 209:504–509.
- Kotrschal K, Sundstro¨m F, Brelin D, Devlin RH, Kolm N (2012). Inside the heads of David and Goliath: environmental effects on brain morphology among wild and growth-enhanced coho salmon *Oncorhynchus kisutch. Journal of Fish Biology*, *81*:987–1002.
- LaDage LD, Maged RM, Forney MV, Roth II TC, Sinervo B, Pravosudov VV (2013). Interaction between territoriality, spatial environment, and hippocampal neurogenesis in male side-blotched lizards. *Behavioral Neuroscience*, *127*:555.
- LaDage L D, Riggs BJ, Sinervo B, Pravosudov VV (2009). Dorsal cortex volume in male side-blotched lizards, *Uta stansburiana*, is associated with different space use strategies. *Animal Behaviour*, 78:91-96.
- Leal-galicia P, Castaneda-bueno M, Quiroz-baez R, Arias C (2008). Neurobiology of Learning and Memory Long-term exposure to environmental enrichment since youth prevents recognition memory decline and increases synaptic plasticity markers in aging. *Neurobiology of learning and memory*, 90:511–518.

- Mendyk RW (2018) Challenging folklore reptile husbandry in zoological parks. In: Berger M, Corbett S (eds) *Zoo animals: husbandry, welfare and public interactions*. Nova Science Publishers, Hauppauge, pp. 265–292.
- Mendyk RW, Augustine L (2023) Controlled deprivation and enrichment. In: Warwick C et al (Eds), *Health and welfare of captive reptiles*, 2nd ed (pp. 323–356). Springer.
- Nagabaskaran G (2020). Factors impacting cognition and cognitive performance in reptiles (Masters Thesis, University of Lincoln).
- Nagabaskaran G; Burman OHP; Hoehfurtner T; Wilkinson A (2021). Environmental enrichment impacts discrimination between familiar and unfamiliar human odours in snakes (*Pantherophis guttata*). *Applied Animal Behaviour Science*, 237:105278
- Nagabaskaran G, Skinner M, Miller N (2022). Western Hognose Snakes (*Heterodon nasicus*) Prefer Environmental Enrichment. *Animals*, 12:3347.
- Näslund J, Aarestrup K, Thomassen ST, Johnsson JI (2012). Early enrichment effects on brain development in hatchery-reared Atlantic salmon (*Salmo salar*): No evidence for a critical period. *Canadian Journal of Fisheries and Aquatic Science*, 69:1481–1490.
- Olsson IAS, Dahlborn K (2002). Improving housing conditions for laboratory mice: a review of environmental enrichment. *Laboratory animals*, *36*:243–270
- Rosier RL, Langkilde T (2011). Does environmental enrichment really matter? A case study using the eastern fence lizard, *Sceloporus undulatus*. *Applied Animal Behaviour Science*, *131*:71–76.
- Phillips CJC, Jiang Z, Hatton AJ, Tribe A, Le Bouar M, Guerlin M, Murray PJ (2011). Environmental enrichment for captive Eastern blue-tongue lizards (*Tiliqua scincoides*). *Animal Welfare*, 20:377-384.
- Scotto Lomassese S, Strambi C, Strambi A, Charpin P, Augier R, Aouane A, Cayre M (2000). Influence of environmental stimulation on neurogenesis in the adult insect brain. *Journal of Neurobiology*, 45:162–171.

- Segovia G, Yagüe AG, García-Verdugo JM, Mora F (2006). Environmental enrichment promotes neurogenesis and changes the extracellular concentrations of glutamate and GABA in the hippocampus of aged rats. *Brain Research Bulletin*, 70:8–14.
- Senn DG (1970). The stratification in the reptilian central nervous system. *Cells Tissues Organs*, 75:521-552.
- Shine R, Charnov EL. (1992). Patterns of survival, growth, and maturation in snakes and lizards. *The American Naturalist*, *139*:1257–1269.
- Spain MS, Fuller G, Allard SM (2020). Effects of habitat modifications on behavioral indicators of welfare for Madagascar giant hognose snakes (*Leioheterodon madagascariensis*). Animal Behavior and Cognition, 7:70-81.
- Spring S, Lerch JP, Henkelman RM (2007). Sexual dimorphism revealed in the structure of the mouse brain using three-dimensional magnetic resonance imaging. *NeuroImage*, *35*:1424–1433.
- Tahamtani FM, Pedersen IJ, Toinon C, Riber AB (2018). Effects of environmental complexity on fearfulness and learning ability in fast growing broiler chickens. *Applied Animal Behaviour Science*, 207:49–56.
- Toli EA, Noreikiene K, DeFaveri J, Merilä J (2017). Environmental enrichment, sexual dimorphism, and brain size in sticklebacks. *Ecology and Evolution*, 7:1691-1698.
- Turner JT, Whittaker AL, McLelland D (2022). Behavioural Impact of Captive Management Changes in Three Species of Testudinidae. *Journal of Zoological and Botanical Gardens*, 3:555-572.
- Van de Weerd HA, Van Loo PLP, Van Zutphen LMF, Koolhaas JM, Baumans V (1996). Preferences for nesting material as environmental enrichment for laboratory mice. *Laboratory Animals*, 31:133-143
- Warwick C (1990). Reptilian ethology in captivity: Observations of some problems and an evaluation if their aetiology. *Applied Animal Behaviour Science*, 26:1–13.
- Warwick C, Steedman C (2023). Naturalistic versus unnaturalistic environments. In *Health and welfare of captive reptiles* (pp. 487-507). Cham: Springer International Publishing.

- Wiedenmayer C (1996). Effect of cage size on the ontogeny of stereotyped behaviour in gerbils. *Applied Animal Behaviour Science*, 47:225-233.
- Wolf SA, Kronenberg G, Lehmann K, Blankenship A, Overall R, Staufenbiel M, Kempermann G (2006).
   Cognitive and physical activity differently modulate disease progression in the amyloid precursor protein (APP)-23 model of Alzheimer's disease. *Biological psychiatry*, 60:1314-1323.
- Young RJ, de Azevedo CS, Cipreste CF (2020). Environmental Enrichment: The creation of opportunities for informal learning. *Zoo animal learning and training*, 101-118.
- Zieliński D (2023). The Effect of Enrichment on Leopard Geckos (*Eublepharis macularius*) Housed in Two Different Maintenance Systems (Rack System vs. Terrarium). *Animals*, 13:1111.

**Acknowledgements:** The authors would like to thank Kelley Putzu for animal care assistance, members of the Collective Cognition Lab for helpful discussions of the results and John Sled and Shoshana Spring for assistance with brain imaging. We also thank two anonymous reviewers for helpful comments on an earlier version of the paper. This research formed part of the work for the PhD of GN. This study was funded by a National Science and Engineering Research Council of Canada (NSERC) Discovery Award to NM (RGPIN-2023-05765).

Statement of interests: The authors declare no conflicting financial or other interests.

**Data availability statement:** All data analyzed in this paper are available in our OSF repository, at: <a href="https://osf.io/ph2bf/?view\_only=7108852060a546bd952bbff57c4935f5">https://osf.io/ph2bf/?view\_only=7108852060a546bd952bbff57c4935f5</a>.